IN VITRO PROTECTIVE EFFECT OF *TELFAIRAI OCCIDENTALIS* (FLUTED PUMPKIN) LEAVES AGAINST OXIDATIVE DNA DAMAGE INDUCED BY REACTIVE OXYGEN SPECIES

BASSA Obed Yakubu*, Abdulkadir Sayyadi, Suleiman Rabiatu Bako, Adejoh Sarah Ufedu, Rilwanu Zainab Julde, Abdulkarim Anisa Garba

Department of Biochemistry, Faculty of Life Sciences, Ahmadu Bello University, Zaria

*Corresponding Author Email Address: <u>obedbassa@gmail.com</u> or <u>obedbassa@abu.edu.ng</u>

Phone: +2348065111379

ABSTRACT

Oxidative DNA damage is an inevitable consequence of cellular metabolism leading to reactive products that cause cellular damage. Telfairai occidentalis is a tropical vine grown in West Africa as a vegetable and for its edible seeds. This study aimed to evaluate the in vitro protective effect of T. occidentalis leaves extract against DNA damage induced by Fenton's system. T. occidentalis leaves were extracted using three solvents; water, nbutanol (n-BuOH) and ethyl acetate (EtOAc). pVax plasmid DNA (476 ng) was incubated with Fenton's system (FeSO₄/H₂O₂) in the presence/varying concentrations of the extract (10 mg/mL, 5 mg/mL and 1 mg/mL) at 37°C for 30 minutes while the control was devoid of extract. The incubate were analysed in 0.8 % agarose gel. The ethyl acetate and the butanolic extract were subjected to GC-MS analysis. The protective effect of T. occidentalis leaves extracts demonstrated an effective protective activity against oxidative stress-induced DNA damage and was dose-dependent with higher dose being most protective. Oleic acid and 3-hydroxy benzoic acid were found to be most likely the bioactive components of the butanolic and the ethyl acetate extract respectively.

Keywords: Reactive Oxygen Species, DNA damage, Telfairai occidentalis

INTRODUCTION

Reactive Oxygen Species (ROS) produced during normal physiological events is a significant contributor to cellular and molecular damage and interacts with biomolecules like lipids, proteins, and nucleic acids, leading to oxidative damage. One of the most concerning effects of ROS is oxidative damage to DNA, which can result in carcinogenesis, mutagenesis and other degenerative diseases such as cancer and neurodegenerative conditions. Mitigating the effects of ROS for biological systems and protecting the DNA from oxidative damage has become an important focus in preventing related diseases (Chaudhary et al

2023). ROS is eliminated by antioxidant defense mechanisms. protecting the cells of its toxic effects have of recent aroused considerable interest due to their broad range of effects in biological systems. Overproduction of ROS in pathological conditions are associated with several chronic disease prevention and development (Kumar and Pandey, 2013, Shen et al. 2022). Naturally occurring antioxidant compounds from plant sources have generated increased attention as potential therapeutic agents for combating ROS and are increasingly investigated for use in foods to replace synthetic antioxidants due to their side effects (Hassanpour and Doroudi, 2023). They are favored for their low toxicity, availability and wide variety of bioactive compounds. T. occidentalis is a tropical vine from West Africa for culinary utilization. It is known for its antioxidant capabilities, but the potential protective effects of the leaves against oxidative damage induced by ROS is yet to be extensively studied. Understanding its protective effect against oxidative DNA damage could provide insight into its potential as a natural antioxidant (Zhuang et al. 2023) and may lead to novel approaches in preventing oxidative stressrelated diseases (Gomaa et al. 2022). We investigate the in vitro antioxidant and protective effect of T. occidentalis leaves against DPPH induced oxidative stress DNA damage.

METHOD

T. occidentalis plant leaf samples were obtained and confirmed at the Herbarium of the department of biological Science, ABU Zaria. Aqueous extract of shade dried, pulverized leaves was fractionated with water, *n*-butanol and ethyl acetate. The IC₅₀ and free radical scavenging activities of *T. occidentalis* on DPPH free radicals was determined according to Oktay *et al.* (2003) with absorbance measured spectrophotometrically at 517nm. The protective effect of *T. occidentalis* on DNA damage was determined using Fenton's system according to Li *et al.* (2013) on *pVax* plasmid DNA (476ng), the outcome was analyzed on 0.8% agarose gel electrophoresis.

Science World Journal Vol. 19(No 4) 2024 www.scienceworldjournal.org ISSN: 1597-6343 (Online), ISSN: 2756-391X (Print) Published by Faculty of Science, Kaduna State University

RESULTS

100

Free radical scavenging activities of T. occidentalis on DPPH free radicals

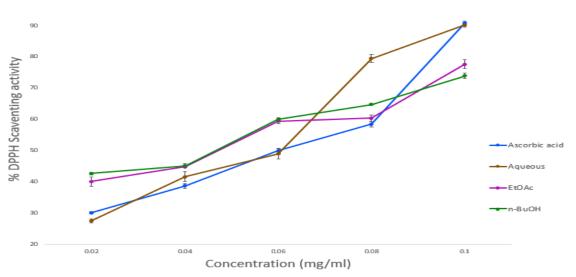
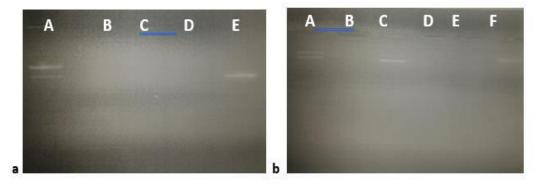


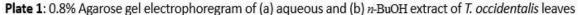
Figure 1: DPPH scavenging activity of the different extracts of T. occidentalis

Figure 1 demonstrated the reduction capability indicating its potential antioxidant activity. IC_{50} for aqueous, ascorbic acid, EtOAc and *n*-BuOH was 50.78, 54.93, 45.77 and 42.38 µg/mL respectively.

radical scavenging activity using DPPH test exhibited significant protection against free radical (DPPH) scavenging activity in a dose dependent pattern, this enhanced the levels of an antioxidant defense system

The potential of *n*-BuOH extract of *T. occidentalis* showed free





- (c) A = Plasmid + 10.0 mg/ml extract + Fenton's, B = Plasmid + 5.0 mg/ml extract + Fenton's, C = Plasmid + 1.0 mg/ml extract D = Plasmid + Fenton's, E = Plasmid + water
- (d) A = Plasmid + extract, B = Plasmid + Fenton's, C = Plasmid + DMSO/PBS (1:1), D = Plasmid + 10.0 mg/ml extract + Fenton's, E = Plasmid + 5.0 mg/ml extract + Fenton's, F = Plasmid + 1.0 mg/ml extract + Fenton's

DNA nicking induced by hydroxyl radical

The pVax plansmid from plate 1a, in Lane A: Plasmid + 10.0 mg/mL + Fenton's reagent showed the pVax plasmid was protected, in lane B: Plasmid + 5.0 mg/mL extract + Fenton's reagent was degraded, in lane C: Plasmid + 1.0 mg/mL + Fenton's reagent was

also degraded, in Iane D: Plansmid + Fenton's (negative control) was degraded and in Iane E: Plasmid + water being the positive control with the reference standard was protected. While the pVax plasmid in plate 1b, in Lane A: Plasmid + extract was protected, Iane B: Plasmid + Fenton's reagent was degraded, Science World Journal Vol. 19(No 4) 2024 www.scienceworldjournal.org ISSN: 1597-6343 (Online), ISSN: 2756-391X (Print) Published by Faculty of Science, Kaduna State University

lane C: Plasmid + DMSO/PBS (1:1) was protected, lane D: Plansmid + 10.0 mg/mL extract + Fenton's reagent was partially protected, lane E: Plasmid + 5.0 mg/mL extract + Fenton's reagent was slightly protected, and lane F: Plasmid + 1.0 mg/mL extract + Fenton's reagent was protected.

The electrophoregram in Plate 1 showed the protection ability of *T* occidentalis (a and b) and reference standard (E) when compared with (D) the negative control was found to be effective in protecting pVax plasmid DNA against the strand breakage induced by hydroxyl radicals in a Fenton's reaction mixture.

DISCUSSION

The current study investigated the protective effect of T. occidentalis leaf extract. The in vitro antiplasmodial activities of T. occidentalis extract demonstrated the leaves to possess significant protective activity, suggesting that T. occidentalis contain biologically active substances. This supports the traditional use of this plant for various medicinal and culinary purposes. T. occidentalis and the reference compound (ascorbic acid) the free radical (DPPH) scavenging activity increased steadily with the increase in concentration. The absorbances at 517 nm of T. occidentalis and ascorbic acid showed that T occidentalis can act as electron donor and can react with free-radicals to convert them to more stable products and thereby terminate radical chain reactions (Ding et al. 2020). The IC₅₀ value of T occidentalis was observed to be at 50.78, 54.93, 45.77 and 42.38 μ g/mL for aqueous, ascorbic acid, EtOAc and n-BuOH was respectively, suggesting a moderately active antplasmodial activity relative to ascorbic acid. According to the norm the extract is active when IC50 < 5 µg/mL, moderately active when 5 µg/mL< IC₅₀ <50 µg/mL and weak when IC50 >50 µg/mL (Berre et al. 2022, Garcia-Molina et al. 2022). The reducing power of plant compounds might be due to the di- and mono-hydroxyl substitution in the aromatic ring which possesses potential hydrogen donating abilities (Ding et al. 2020). The reducing properties are generally associated with the presence of reductones, known to exert antioxidant activity by breaking the free-radical chain by donating a hydrogen atom (Konecny et al. 2018, Savel et al. 2012).

Hydroxyl radical is the most reactive among reactive oxygen species (Behnaz et al. 2022), and considered to be responsible for much of the biological damage in free-radical pathology (Mittal et al. 2023), has the capacity to cause strand breakage in DNA, and contributes to carcinogenesis, mutagenesis and cytotoxicity (Chaudhary et al 2023). The DNA protective effect of aqueous, n-BuOH and EtACc extract of T. occidentalis leaf was checked against Fenton's induced DNA damage on pVax plasmid DNA. The protection offered against DNA damage by T. occidentalis was concentration dependent at concentration 0.02-0.1mg/mL, protection exhibited was more effective and slightly close to that of ascorbic acid. ROS however play a two-fold role as both toxic (generating oxidative stress) and beneficial (indulge in cellular signaling and immune function) compounds to the organism's system (Chaudhary et al 2023). The result showed a correlation between the effect and concentration, and attributed to the presence of high amounts of polyphenols, a potent antioxidants

Conclusion

T. occidentalis extracts exhibited an effective ability to scavenge DPPH radicals in a dose-dependent manner. It was found to be

effective in protecting plasmid DNA against damage induced by hydroxide radicals in Fenton's reagent and can be a rich source of antioxidants that mitigate oxidative stresses on susceptible biomolecules.

REFERENCES

- Behnaz Akbari, Baghael-Tazdi Namdar, Manochehr Bahmale, Fatemeh Mahdavi Abhari (2022). The role of plant-derived natural antioxidants in reduction of oxidative stress. Journal of BioFactors. Vol 48, issue 3, doi: 10.1002/biof.1831
- Berre Le M., Gerlach Jared Q., Dziembala Iwona, Kilcoyne M. (2022). Reactive oxygen species in inflammation and tissue injury. *Antioxidant and Redox* 20(7) doi: 10.89/ars.20125149
- Chaudhary P., Janmeda Pracheta, Docea A.m Yeskaliyeva B., Abdull Razis A., Modu B., Calina D., Sharifi-Rad Javad (2023) Oxidative stress, free radicals and antioxidants: potential cerosstalk in the pathophysiology of human diseases. *Frontiers in Chemistry* 11 (2023) doi: 10.3389/fchem.2023.1158198
- Ding Xiaochu, Chen Ying, Chao A. C., Wu Yen-Lin, Wang Yadong (2020) Control the Mechanical Properties and degradation of Poly (glycerol sebacate) by substitution of the hydroxyl groups with palmitates. *Macromolecular Bioscience* 20(9) doi: 10.1002/mabi.202000101
- Garcia-Molina P., Teruel-Puche J., Rodriguez-Lopez N., Garcia-Canovas F., Monoz-Munoz J. (2022). The relationship between the IC50 values and the apparent inhibition constant in the study of inhibitors Tyrosinase Diphenolase Activity helps confirm the Mechanism of inhibition. *Molecules* 27(10) Doi: 10.3390/molecules27103141
- Hassanpour S. Doroudi Alireza (2023). Review of the antioxidant potential of flavonoids as a subgroup of polyphenols and partial substitute for synthetic antioxidants Avicenna Journal of Phytomedicine 13(4) doi: 10.22038/AJP.2023.21774
- Konecny M., Cejpek K., Cechovska L., Velisek J., (2018) Transformation pathways of redcutones in the advanced maillard reaction. Journal of Food Sciences 27(10):5149-5152 doi: 10.17221/1091-CJFS
- Kumar Shashank and Pandey Abhay K. (2013). Chemistry and Biological Activities of Flavonoids: An Overview. The Scientific World Journal Vol. 2013, 1-16 doi.org/10.1155/2013/162750
- Lee J. C., Kim, H. R., Kim, J. and Jang, Y. S. (2002). Antioxidant property of an ethanol extract of the stem of *Opuntia ficusindica* var. saboten. Journal of Agricultural and Food Chemistry, 50(22): 6490–6496. Doi: 10.1021/jf020388c
- Mittal Manish, Mohammad Rizwan Siddiui, Khiem Tran, Sekhar P. Reddy, Asrar B. Malik (2014). Reactive Oxygen species in inflammation and tissue injury Antioxidant redox Signal 1:20(7):1126-67. Doi: 10.1089/ars.2012.5149.
- Oktay Munir, Gulcin Ilhami, Irfan O. Kufreviioglu (2003). Determination of *in vitro* antioxidant activity of fennel (Feoniculum vulgare) seed extracts. 36(2), 0-271. Doi: 10.1016/s0023-6438(02)00226-8
- Ozaki T, Nakamura M, and Shimozato O. (2015) Novel implications of DNA damage response in drug resistance of malignant cancers obtained from the functional interaction between p53 family and RUNX2 Biomol. 5:2854-2876
- Savel J., Kosin P., Broz A., (2012). Redox power changes of caramels and sugar reductones in beer. *Journal of Field Rebotics* 1(1) doi: 10.5539/JFR.V1N1P132